

High-throughput gene function annotation using transposon mutagenesis

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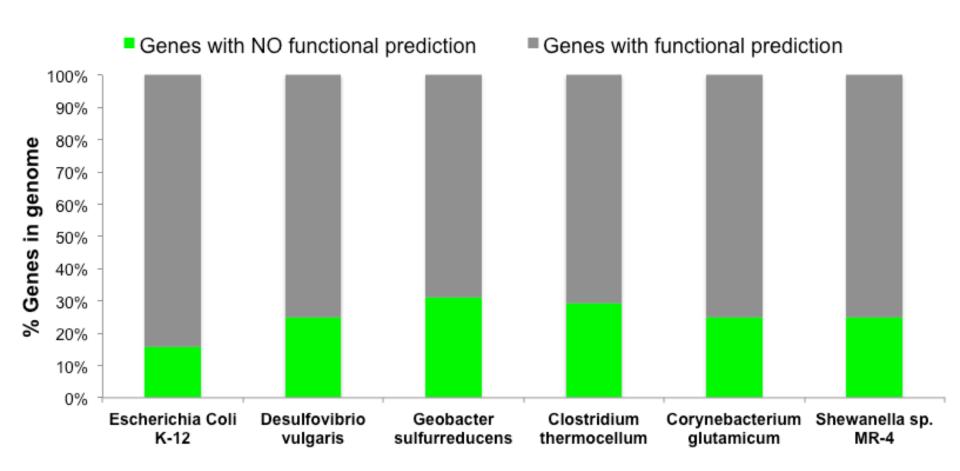
Outline



- Overview of transposon mutagenesis and sequencing (TnSeq)
- Applications and examples
- Development of RB-TnSeq for high-throughput gene function annotation in diverse bacteria

Microbial genomes are poorly characterized





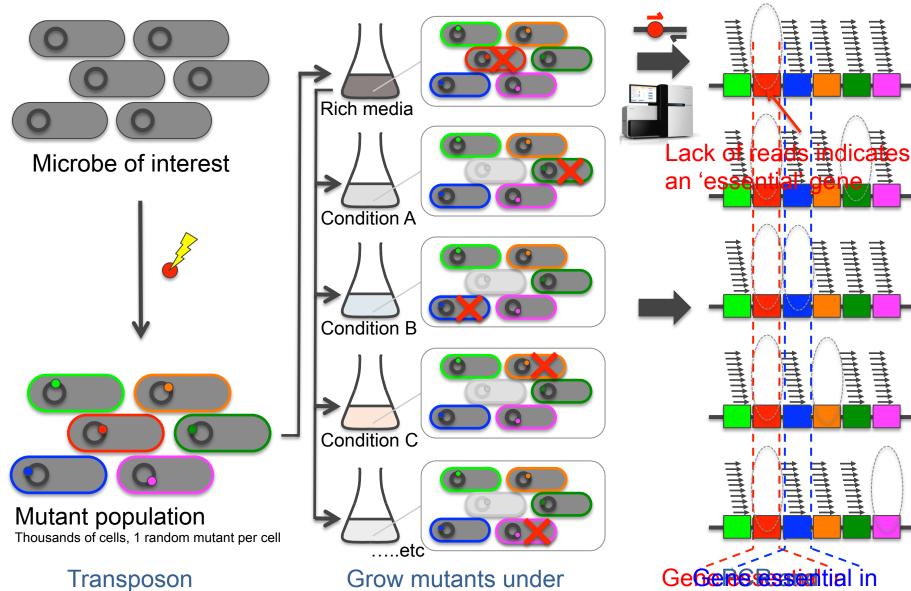
Up to 30% genes in microbial genomes have no known function

Gene function annotation by transposon mutagenesis and sequencing (TnSeq)

Mutagenesis



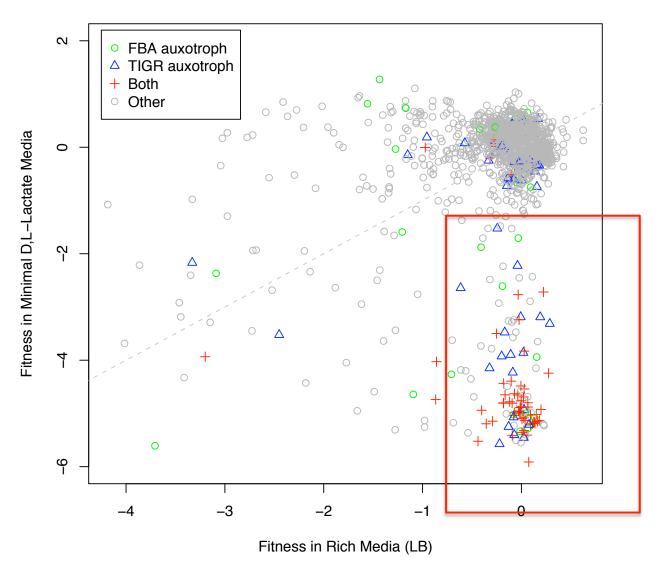
all conditions being



many conditions

Fitness in rich versus minimal media



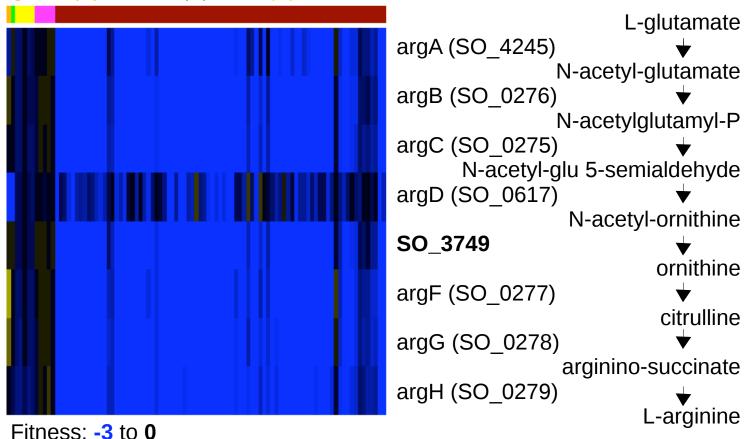


Fill holes in biosynthetic pathways



<u>Carbon/Nitrogen sources:</u>

Arginine (N), Gelatin (C), CAS (C), LB, Other defined

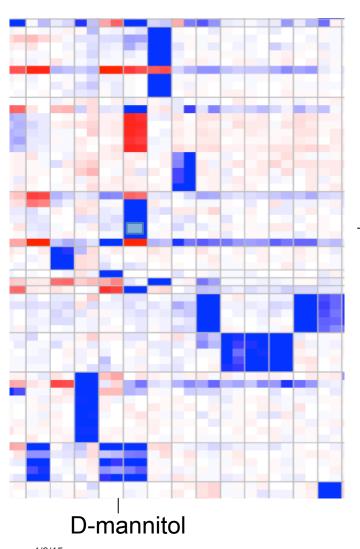


Example from Shewanella oneidensis MR-1

Fill holes in catabolic pathways



Phaeobacter inhibens BS107



Gene Fitness

-3 0 3
gene is gene is
important detrimental

PGA1_c16670

"putative N-acylglucosamine 2-epimerase"

Actually, D-mannose isomerase

Predicted for *Shewanella amazonensis* (Rodionov et al. 2010) and also confirmed by our data.

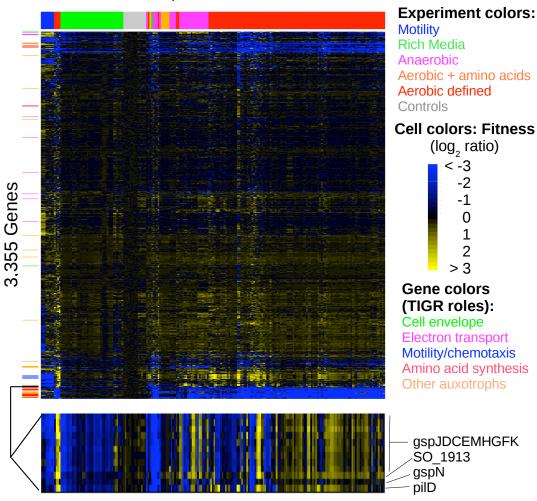
Also identified roles for transporters and regulators

Global annotation of gene function



Example from Shewanella oneidensis MR-1

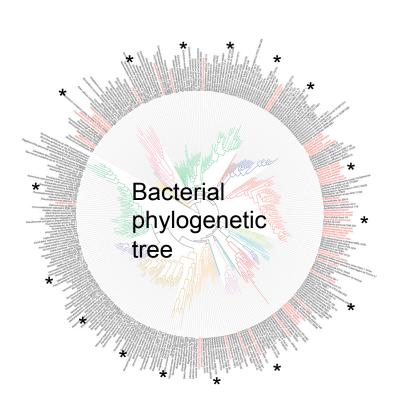
195 Experiments & 19 Controls

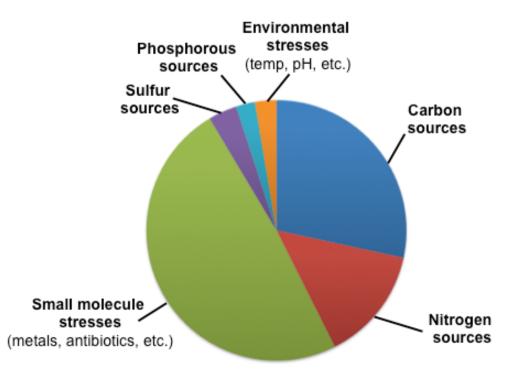


- Phenotypes for over 2,000 genes including hundreds of previously uncharacterized genes
- Used fitness data to infer functions for 41 genes or operons

Functional Encyclopedia of Bacteria and Archaea (FEBA)







Mutagenesis of 20 diverse organisms

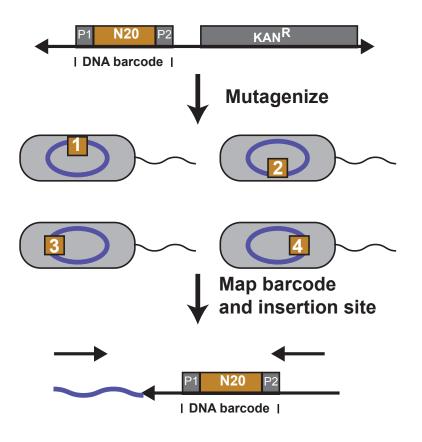


~100 growth conditions

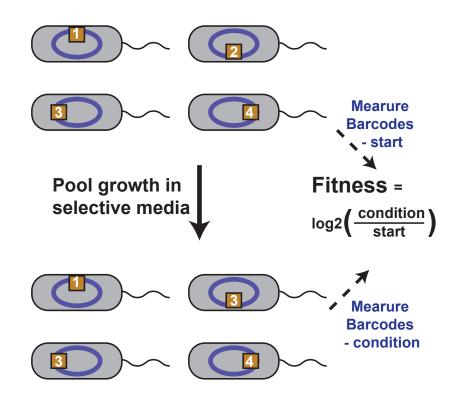
Random barcode transposon site sequencing (RB-TnSeq)



A. Characterize mutant population



B. Assay mutant fitness with DNA barcodes



See Wednesday poster - Kelly Wetmore

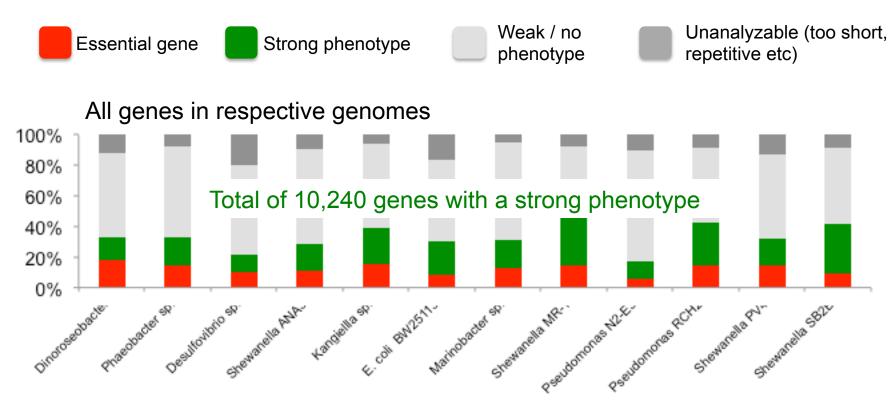
Properties of 20 FEBA mutant libraries



Phylum / Class	Species	# Mutants
alpha-proteobacteria	Phaeobacter inhibens	215,858
	Dinoroseobacter shibae	119,703
	Sphingomonas koreensis	226,420
beta-proteobacteria	Azospira suillum PS	241,691
	Cupriavidus sp. 4G11	112,124
	Marinobacter adhaerens	85,718
	Proudomonas en N2 E3	510 225
Average 170,000 mutants per genome Average 60 mutants per gene		
gamma-proteobacteria	Shewanella oneidensis	181,569
	Pseudomonas sp. N1-B4	176,258
	Pseudomonas stutzeri RCH2	166,448
	Escherichia coli	152,018
	Pseudomonas sp. N2-E2	110,065
	Kangiella aquimarina	100,069
	Shewanella sp. ANA3	69,557
	Shewanella sp. PV4	51,094
delta-proteobacteria	Desulfovibrio miyazaki	28,252
cyanobacteria	Synechococcus elongatus	141,062

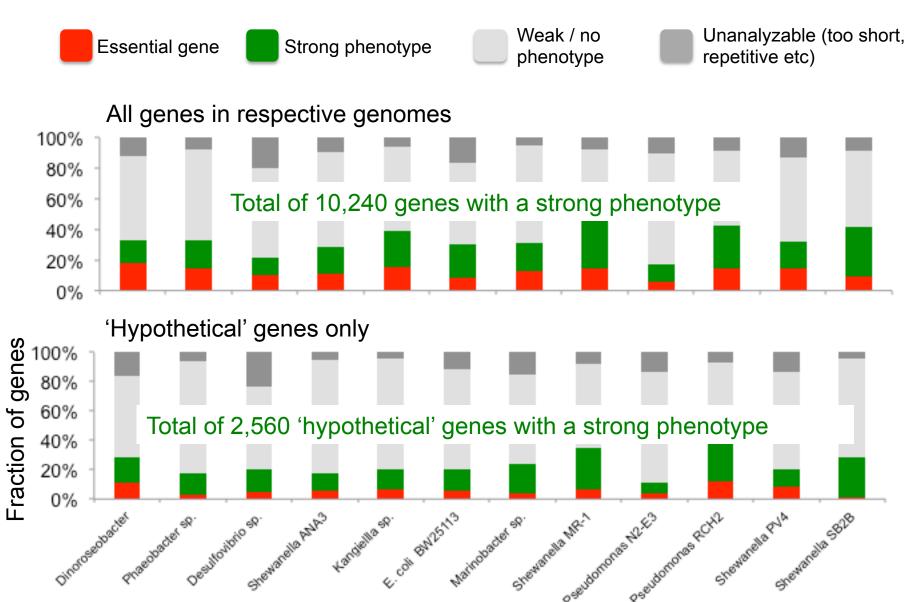
Summary of results from 12 bacteria





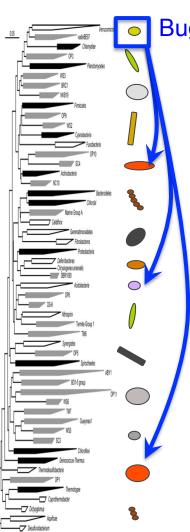
Summary of results from 12 bacteria





Can mutant fitness annotations be propagated across diverse genomes?



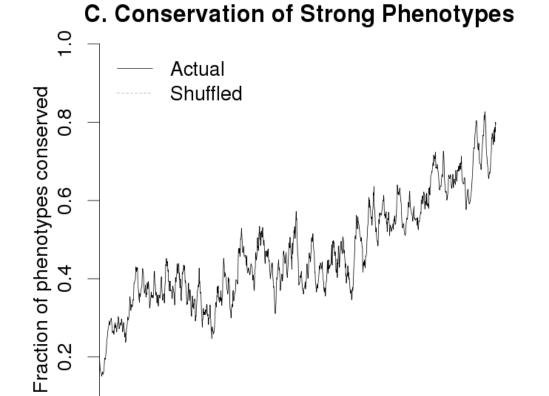


Bug with FEBA annotations

Does: sequence conservation = function conservation ?

Strong phenotypes are often conserved between distant homologs





0.6

Similarity to putative ortholog (score ratio)

0.2

0.4

Shows why annotation by homology (mostly) works

Can objectively define "functional orthologs" to study evolution?

4/8/15

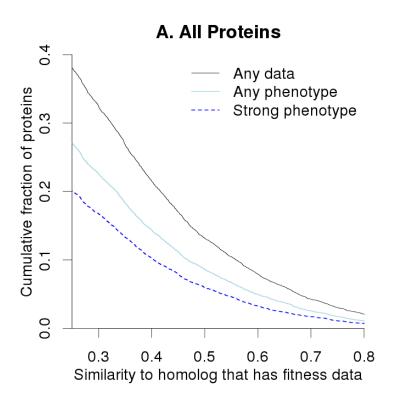
1.0

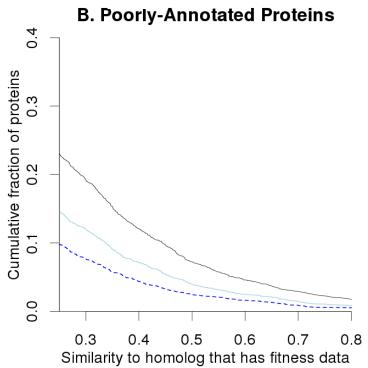
0.8

Coverage of all bacterial proteins



Take a random sample of bacterial proteins: do we have phenotypes for their homologs?

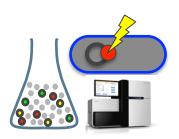




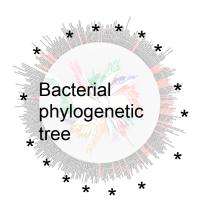
$$similarity(A, B) = \frac{protein BLAST score}{self score}$$

Summary





We have developed RB-TnSeq, a highthroughput approach to annotate gene function in bacteria



The 'FEBA' project will provide functional annotation for 20 diverse organisms / 1000s novel genes



Future 'product' of JGI? Keen to target bugs of interest to DOE and to JGI user community

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