

Oxford Nanopore Sequencing

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Outline



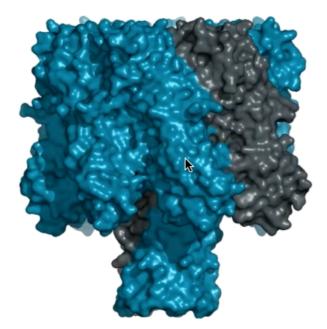
- Technology Introduction
- Current Experience:

Overview of ONT library types

- Sequencing Run Performance
- Application: Mini Mock Metagenome
- Future Directions

Nanopore Sequencing

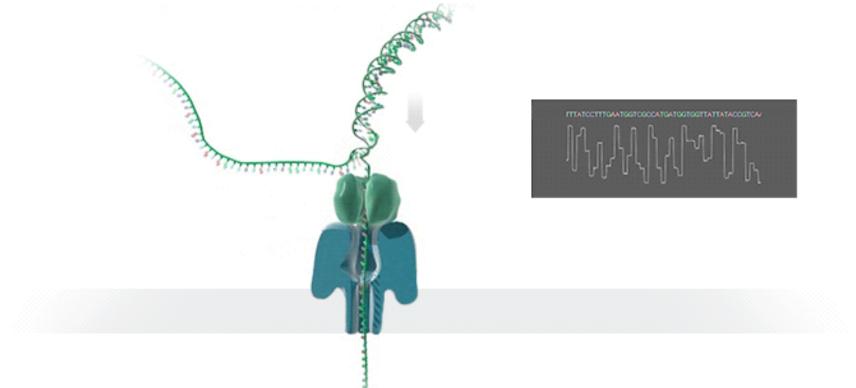




Source: Oxford Nanopore Technologies

ONT Sequencing

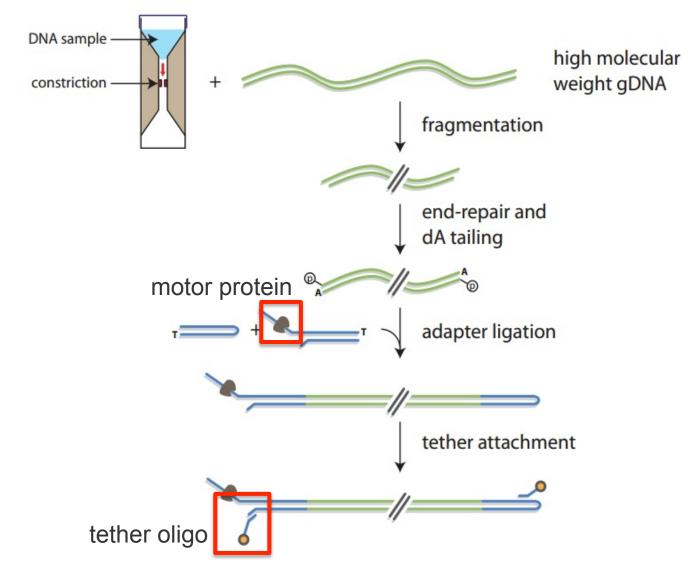




No polymerase or amplification needed \rightarrow as long as DNA intact, long reads possible

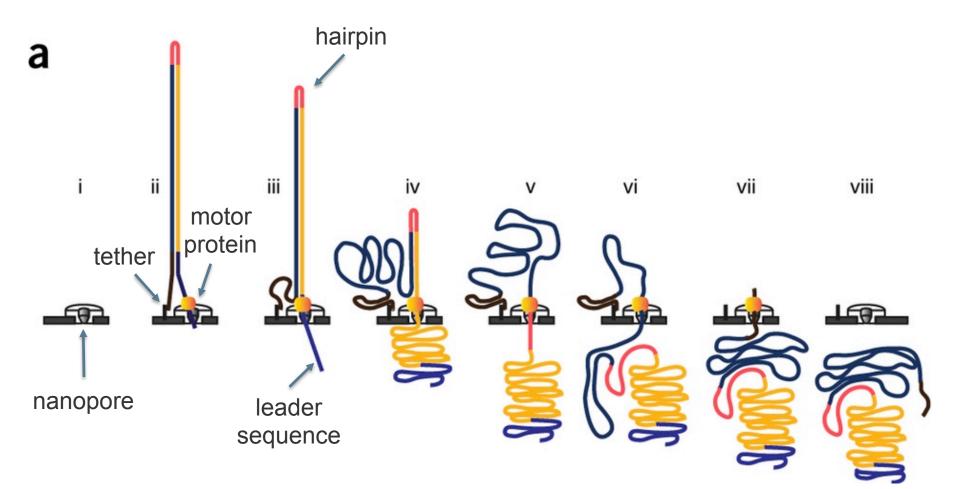
Simple Library Preparation





Nanopore Sequencing

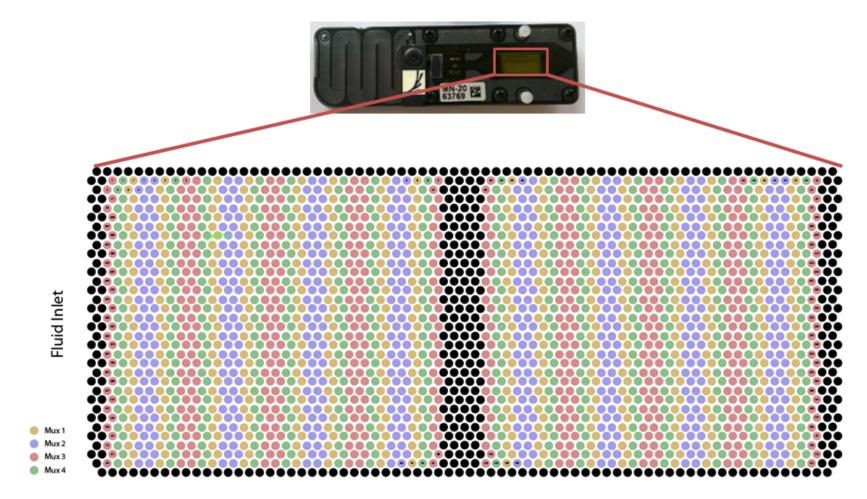




Source: Jain et al., Nature Methods 2015

Flow Cell Layout



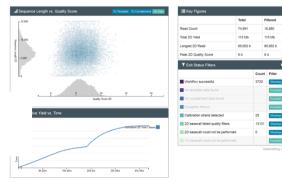


512 sequencing channels \rightarrow each channel sequences multiple molecules

Cloud-Based Data Processing















Outline



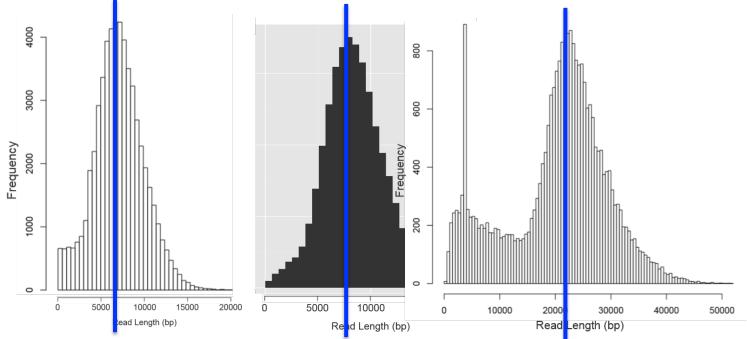
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Library Types

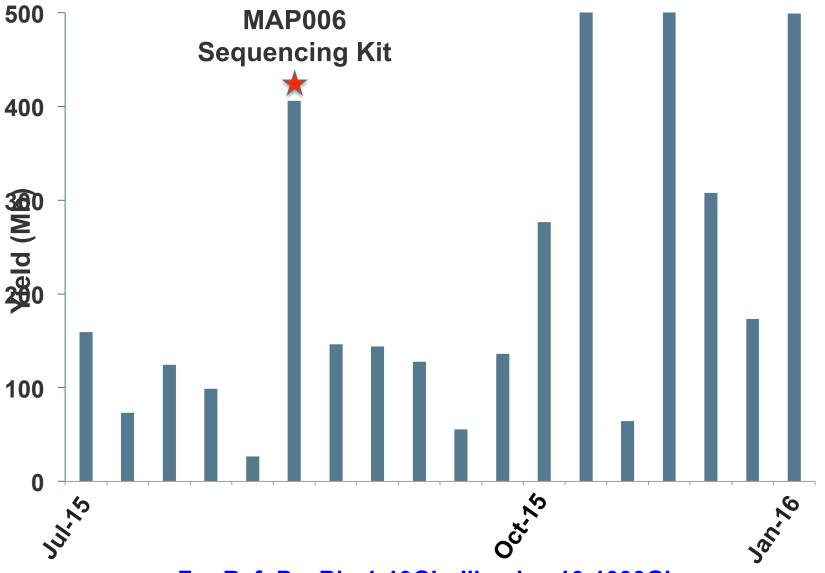




Run Statistic	8kb Standard	8kb Low Input	20kb
Input	1µg	100ng	15µg
Yield (Mb)	406 Mb	303 Mb	537 Mb
# Reads	57K	37K	26K
Mean Size	7263 bp	8524 bp	21310 bp

Review of Runs – Yield (Mb)



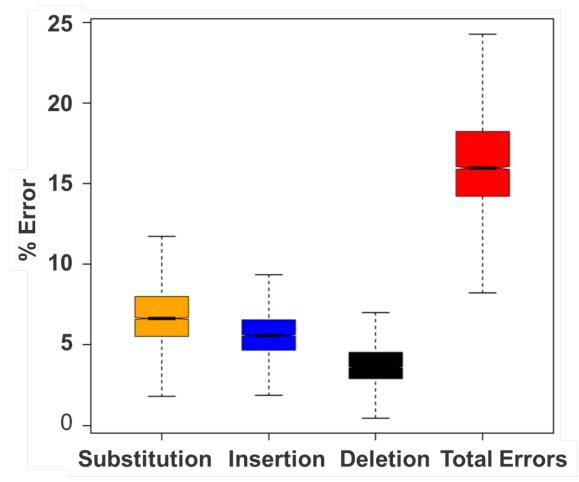


For Ref: PacBio 1-10Gb; Illumina 10-1000Gb

High Error Rate



- **Percent ID = ~ 83%**
 - Note: Illumina >99%, PacBio ~ 85%
- Substitutions > Insertions > Deletions



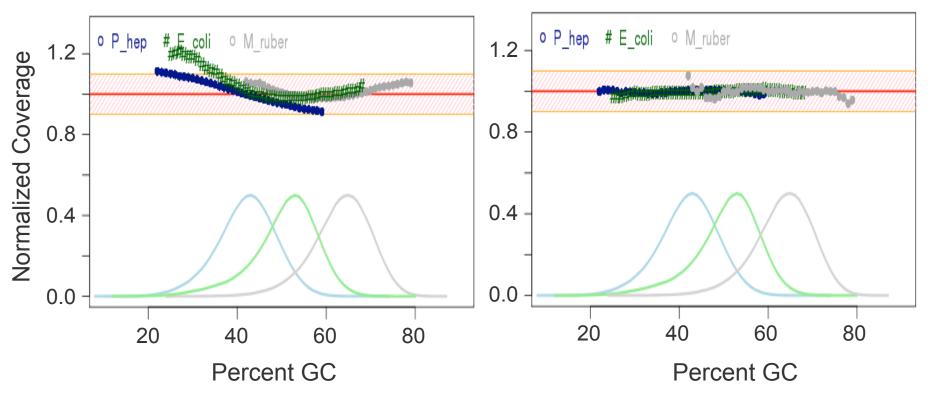
ONT Minimal GC Coverage Bias



Species	P. heparinus	E. coli	M. ruber
GC content	42%	51%	63%

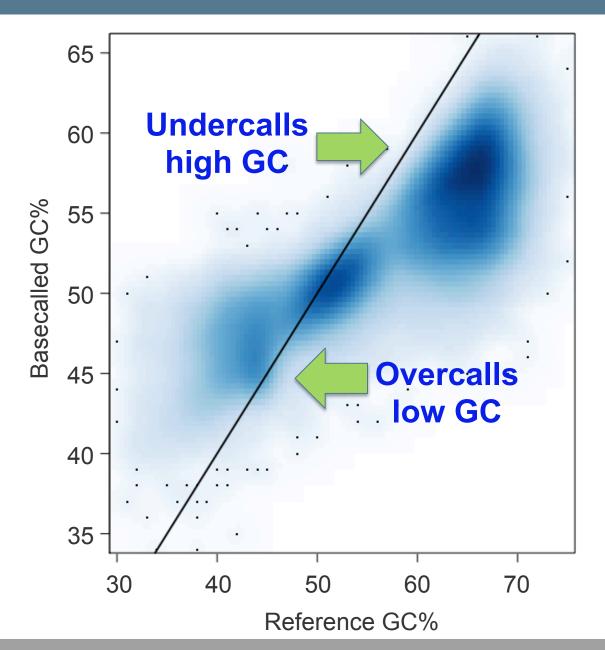
Illumina

ONT



Incorrect GC Calls at Extremes

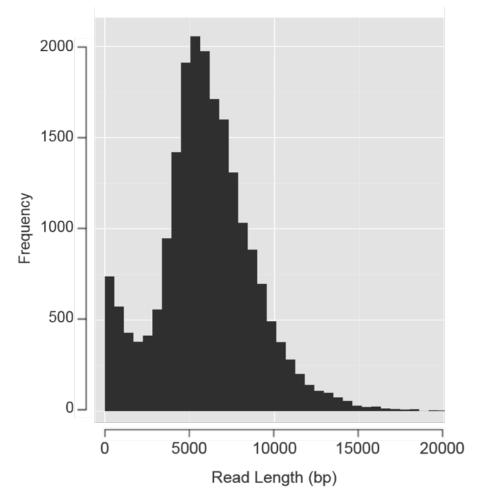




Application: Mini Mock Metagenome



- 4 organisms pooled in equimolar amounts
 - GC content from 40% to 70% -- 2 organisms with GC content > 60%
- 8 kb library constructed and sequenced



Run Statistic	Amount
Yield (Mb)	458 Mb
# Reads	77K
Mean Size	7723 bp
Median Size	7674 bp

ONT: Conclusions and Considerations



- 8kb Protocol Modifications Yield 20kb+ Libraries
 - Consistent and robust protocol performance
- 100ng Low-Input Protocol Provides Similar Yields
- Yields and Read Lengths from Mini Mock Metagenome Similar to Single Organisms
- Currently Higher Error Rate in Extreme GC Content
- Yet, Minimal GC Coverage Bias Observed

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Future Directions



• Protocol development and applications:

- Ex. longer read lengths
- Ex. complex metagenome communities

• New ONT protocols:

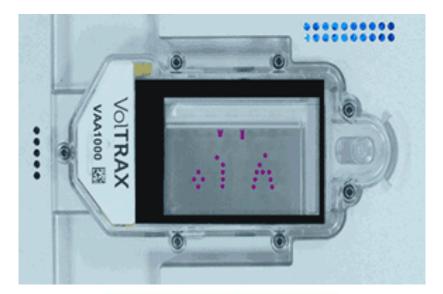
- New sequencing chemistry (R9) faster speed, more accurate basecaller
- Sequencing of native RNA

• Software development:

- minoTour analysis platform
 - Monitor reads in each pore in real-time and reject reads that are unacceptable

ONT Updates -- Hardware





VOLTRAX

- Automated sample preparation
- 2016



PROMETHION

- 300X greater throughput than MinION
- JGI early access site
- 2016

Director's Call for Proposals



- Use ONT's long-read sequencing capability for novel applications
- http://jgi.doe.gov/user-program-info/other-programs/

Thanks to the JGI Nanopore Group

Chia-Lin Wei Len Pennacchio

JGI

Chee-Hong Wong Chris Daum Denis Tolkunov Matt Zane Rob Egan Volkan Sevim